

Isolation of Microsatellite Markers for the Red Mangrove, *Rhizophora mangle* (Rhizophoraceae)

Author(s): Diana O. Ribeiro, Christina C. Vinson, Dulcivania S. S. Nascimento, Ulf Mehlig, Moirah P. M. Menezes, Iracilda Sampaio, and Marivana B. Silva

Source: Applications in Plant Sciences, 1(9)

Published By: Botanical Society of America

<https://doi.org/10.3732/apps.1300003>

URL: <http://www.bioone.org/doi/full/10.3732/apps.1300003>

BioOne (www.bioone.org) is a nonprofit, online aggregation of core research in the biological, ecological, and environmental sciences. BioOne provides a sustainable online platform for over 170 journals and books published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Web site, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/page/terms_of_use.

Usage of BioOne content is strictly limited to personal, educational, and non-commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

ISOLATION OF MICROSATELLITE MARKERS FOR THE RED MANGROVE, *RHIZOPHORA MANGLE* (RHIZOPHORACEAE)¹

DIANA O. RIBEIRO², CHRISTINA C. VINSON³, DULCIVANIA S. S. NASCIMENTO², ULF MEHLIG²,
MOIRAH P. M. MENEZES², IRACILDA SAMPAIO², AND MARIVANA B. SILVA^{2,4}

²Institute for Coastal Research, Universidade Federal do Pará (UFPA), Alameda Leandro Ribeiro s/n, Aldeia, 68600-000 Bragança, Pará, Brazil; and ³Molecular Biology and Biochemistry, Universidade Federal de Viçosa, Viçosa, 36570-000 Minas Gerais, Brazil

- *Premise of the study:* Three species of the mangrove tree genus *Rhizophora* are found in the New World and along the west coast of Africa. Of these, *R. mangle* is the most abundant and has a complex interbreeding relationship with the sympatric *R. racemosa* and *R. harrisonii*. The development of additional microsatellite markers would permit paternity analyses and investigation of the hybrid origin of these species.
- *Methods and Results:* Using an enriched library method, via hybridization with biotinylated oligonucleotides complementary to repetitive poly AG/TC, primers for 11 microsatellite markers of *R. mangle* were developed and characterized in populations in Pará and São Paulo (Brazil) and Florida (USA). Ten of these markers were transferable to *R. racemosa* and *R. harrisonii*.
- *Conclusions:* The microsatellite markers presented here will be useful in studies of contemporary and historical gene flow between American and West African *Rhizophora* species.

Key words: mangrove; microsatellites; *Rhizophora harrisonii*; *Rhizophora racemosa*; Rhizophoraceae.

In the New World and along the west coast of Africa, three species of the mangrove tree genus *Rhizophora* L. (Rhizophoraceae) have been identified. Of these, *R. mangle* L. (red mangrove) is the most widely distributed (Tomlinson, 1986). In terms of interbreeding, *R. mangle* has a complex relationship with the sympatric *R. racemosa* G. Mey. and *R. harrisonii* Leechm. (Céron-Souza et al., 2010), which could potentially be resolved by the use of microsatellite markers. Two studies have previously reported the isolation of microsatellite markers for *R. mangle*. In Rosero-Galindo et al. (2002), 10 polymorphic loci with between two and seven alleles per locus were reported (26 alleles total). In Takayama et al. (2008), 14 simple sequence repeat (SSR) loci were reported and 11 were shown to be polymorphic, with a total of 32 and 27 alleles in populations from the Pacific and Atlantic coasts of Costa Rica, respectively. The allelic diversity reported for previously isolated microsatellite markers for this species is therefore somewhat low, and this is also the case in subsequent studies that made use of these markers. Arbeláez-Cortes et al. (2007) report a total of 17 alleles for three markers in five studied populations, Pil et al. (2011) report 22 alleles for eight markers in 10 populations, and Sandoval-Castro et al. (2012) report 19 alleles for six markers in 10 populations. For this reason, we isolated and characterized

new microsatellite markers for *R. mangle*, which were then subsequently tested for transferability to *R. racemosa* and *R. harrisonii*. These markers present higher allelic diversity than has previously been reported. In conjunction with pre-existing markers, these new markers will permit mating systems analysis, paternity tests, and future studies attempting to resolve the potential hybrid origin of *Rhizophora* species.

METHODS AND RESULTS

Genomic DNA was isolated from leaf tissue of one adult individual of *R. mangle* sampled in the Caeté estuary, Bragança, Pará, northern Brazil (0°52'22"S, 46°39'04"W), and used in the development of the enriched library. Herbarium material was deposited at the herbarium of the Institute for Coastal Research in Bragança (HBRA; accession no. 821). Polymorphism was evaluated using the 24 adult trees of *R. mangle* from Pará, Brazil (0°50'27"S, 46°38'15"W; HBRA 706); São Paulo, Brazil (24°00'21"S, 46°17'59"W); and Florida, USA (25°08'09"N, 80°57'29"W; HBRA 1291). The São Paulo and Florida sites are near the southernmost and northernmost distributional limits of *R. mangle*, respectively. For the transferability test, we genotyped seven individuals of *R. harrisonii* and five individuals of *R. racemosa*.

An enriched library was constructed according to Vinson et al. (2005). Total DNA (50,000 ng) was digested with *Sau3I* and separated on a 2% agarose gel. Fragments of 300–800 bp were recovered with purification of the gel using QIAquick Gel Extraction kit (QIAGEN, São Paulo, Brazil). DNA fragments were ligated to adapters, hybridized to biotinylated (AG)₁₃ and (TC)₁₃, and the oligofragments separated using streptavidin magnetic beads. Selected fragments were ligated into a pGEM-T Easy Vector (Promega Corporation, Madison, Wisconsin, USA) and transformed in competent *E. coli* XL1-Blue cells that were grown overnight on 1× Luria–Bertani agar plates containing ampicillin, X-gal, and IPTG. Transformants (white colonies) were diluted in water, and the fragment inserted into the plasmid was amplified using the M13 universal primers (M13F[–20] and M13R[–40]) by PCR. PCR products were sequenced on a MegaBACE 1000 instrument (GE Healthcare, Belo Horizonte, Minas Gerais, Brazil) using dye terminator fluorescent chemistry (Sanger method) with the

¹Manuscript received 4 January 2013; revision accepted 15 April 2013.

The authors thank T. C. R. Williams for revising the text of the manuscript and the anonymous reviewers for their helpful comments. D.O.R. received a Masters scholarship from the Fundação de Amparo à Pesquisa do Estado do Pará (FAPESPA).

⁴Author for correspondence: marivana@ufpa.br

TABLE 1. Characteristics of 11 microsatellite loci developed in *Rhizophora mangle*.

Loci	Primer sequences (5′–3′)	Repeat motif	Allele size (bp)	T _a (°C)	GenBank accession no.
RmBra18	F: TCAAGGACAGGTCAACAGCA R: TGAATTAAGACGTCAAATCATCG	(TC) ₁₈	174	54	HQ850264
RmBra19	F: GAGGCAGAGTCAGGTCAGAA R: CACTGGTCCACTGACAGCAA	(CT) ₁₄	162	54	HQ850265
RmBra20	F: TCAGCACAATACATCAGGACAA R: GGCCTATCCATCCTGAGT	(AG) ₁₆	141–175	50	HQ850255
RmBra25	F: TCCATTCTGCCAAATTTGATT R: CGAATTCACCGATCACAATG	(AG) ₁₃	172–226	50	HQ850256
RmBra27	F: TCTTTTGTGCGAAGCCTCTCC R: ATCAAGTTGGCGTTGGAAAA	(CT) ₅ (TC) ₆ (CT) ₄ (TC) ₃	140–184	50	HQ850257
RmBra45	F: GAAAATGCAAGAGGGCTGAC R: CATGGGTCTCTCTCTCTCTGAC	(AG) ₁₇ (AGG) ₃	128–164	54	HQ850258
RmBra50	F: ATCGTGGGAAGAAGCGGGTTF R: TCAAGAAGTCCAGGGTGCTT	(AG) ₁₈	149–215	50	HQ850259
RmBra59	F: GTGAACGCTCTGGACTGGAG R: TCACCGATCCCCTAGAACTG	(AG) ₂₄	145–209	54	HQ850260
RmBra64	F: CAAGGCTAATCCGAAAATCG R: ACTCTGTCCCCTGCCTCAC	(AG) ₁₅	148	54	HQ850266
RmBra65	F: CCAATAGACAATACAGGATACCCAGA R: CAGCATGAACATCACCTTGG	(AG) ₁₂	234	52	HQ850263
RmBra66	F: TCAGCACAATACATCAGGACAA R: GCGCTATCCATCTGAGTTT	(AG) ₁₉	138–206	54	HQ850261

Note: T_a = annealing temperature.

M13F(–20) primer. The sequences were subsequently edited manually using Sequencher version 4.1.4 software (Gene Codes Corporation, Ann Arbor, Michigan, USA). Primers were designed in the regions flanking AG/CT repeats using Primer3 OUTPUT (Rozen and Skaletsky, 2000) with the following conditions: final amplicon between 100 and 350 bp, average annealing temperature of 55°C, primer length between 18 and 22 bp, and GC content between 40% and 60%. We sequenced 66 clones, and 23 pairs of primers were synthesized. The preliminary polymorphism test was conducted with 12 individuals from the Caeté estuary (Bragança). PCR products were visually compared with the 10-bp ladder in a 10% polyacrylamide gel stained with ethidium bromide.

From the 23 primers, 11 primers produced PCR products, seven loci were polymorphic, and four loci were monomorphic (RmBra18, RmBra19, RmBra64, and RmBra65; Table 1). Forward primers of the seven loci were fluorescently labeled with 6-FAM (RmBra20, RmBra25, and RmBra59; MWG-Biotech, Ebersberg, Germany) or HEX (RmBra27, RmBra45, RmBra50, and RmBra66; MWG-Biotech). Loci were amplified using PCR in a total volume of 13 µL containing: DNA (5 ng), 1× PCR reaction buffer (10 mM Tris-HCl [pH 8.3], 50 mM KCl, 1.5 mM MgCl₂), forward primer and reverse primer (0.15 µM), MgCl₂ (1.5 mM), bovine serum albumin (BSA; New England Biolabs, Hitchin, Hertfordshire, United Kingdom; 0.75 mg·mL⁻¹), dNTP (0.5 mM), and 1.3 U *Taq* polymerase (Invitrogen, Life Technologies, Carlsbad, California, USA). PCR conditions were: denaturation at 96°C for 2 min; 30 cycles of denaturation at 94°C for 15 s, annealing at 50–54°C (Table 1) for 15 s, and extension at 72°C for 15 s; and a final extension at 72°C for 5 min. Transfer of the primers of *R. mangle* to *R. racemosa* and *R. harrisonii* was tested under the same conditions as for *R. mangle* (described above). Fragments were genotyped on a 96-capillary sequencer MegaBACE 1000 (GE Healthcare). Fragments were analyzed using MegaBACE Fragment Profiler version 1.2 (Amersham Bioscience, Belo Horizonte, Minas Gerais, Brazil) using MegaBACE ET 550-R Rox Size Standard (Amersham Bioscience). Number of alleles per locus, observed and expected heterozygosities, coefficient of fixation index, and Hardy–Weinberg and linkage disequilibrium were calculated using FSTAT (Goudet, 2002). Probabilities of paternity exclusion were estimated using CERVUS (Kalinowski et al., 2007). Frequency of null alleles was estimated using MICRO-CHECKER version 2.2.3 (van Oosterhout et al., 2004).

Results show a total of 90 alleles in the three studied populations (49 in Bragança, 39 in São Paulo, and 42 in Florida), and the number of private alleles varied between one to eight alleles per locus (Table 2). Observed heterozygosity was lower than values of heterozygosity under expected Hardy–Weinberg equilibrium (HWE) for all populations tested, with higher fixation index values for the São Paulo and Florida populations (Table 2). The test of pairwise linkage after the Bonferroni correction showed values higher than the significant value of 0.005, indicating that these loci are not linked and can be used as independent markers. Only one locus, RmBra27, showed significant null alleles using

the Oosterhout index (0.26). Fisher’s exact test revealed deviations from HWE after Bonferroni correction at two loci for all populations tested, with an excess of homozygotes for the loci RmBra25, RmBra27, and RmBra50 ($P = 0.019$). The combined exclusion power of all seven loci was 0.998, 0.998, and 0.982 for first parent total exclusion probabilities for the Bragança, São Paulo, and Florida populations, respectively. Transfer of the primers was successful, with six polymorphic markers identified for *R. mangle* also polymorphic in *R. racemosa* and *R. harrisonii* (except RmBra25), while the four monomorphic markers identified for *R. mangle* were only tested for amplification.

CONCLUSIONS

The high number of private alleles in the studied populations indicates that these loci are useful for studies of genetic divergence, genetic structure, and phylogeography. In addition, the parental exclusion probability values are adequate for progeny analysis and paternity tests for further identification of reproduction barriers and identification of hybridization between species of all American and West African *Rhizophora* species.

LITERATURE CITED

- ARBELÁEZ-CORTES, E., M. F. CASTILLO-CÁRDENAS, N. TORO-PEREA, AND H. CÁRDENAS-HENAO. 2007. Genetic structure of the red mangrove (*Rhizophora mangle* L.) on the Colombian Pacific detected by microsatellite molecular markers. *Hydrobiologia* 583: 321–330.
- CERÓN-SOUZA, I., E. RIVERA-OCASIO, E. MEDINA, J. A. JIMÉNEZ, W. O. McMILLAN, AND E. BERMINGHAM. 2010. Hybridization and introgression in New World red mangroves *Rhizophora* (Rhizophoraceae). *American Journal of Botany* 97: 945–957.
- GOUDET, J. 2002. FSTAT: A program to estimate and test gene diversities and fixation indices. Website <http://www2.unil.ch/popgen/softwares/fstat.htm> [accessed February 2002].
- KALINOWSKI, S. T., M. L. TAPER, AND T. C. MARSHALL. 2007. Revising how the computer program CERVUS accommodates genotyping error increases success in paternity assignment. *Molecular Ecology* 16: 1099–1106.
- PIL, M. W., M. R. T. BOEGER, V. C. MUSCHNER, M. R. PIE, A. OSTRENSKY, AND W. A. BOEGER. 2011. Postglacial north–south expansion of populations of *Rhizophora mangle* (Rhizophoraceae) along the Brazilian

TABLE 2. Results of primer screening for seven polymorphic microsatellite loci in three populations of *Rhizophora mangle*.

Loci	Bragança						São Paulo						Florida						
	N	A	A _p	H _e	H _o	P _{ex}	N	A	A _p	H _e	H _o	P _{ex}	N	A	A _p	H _e	H _o	F _{IS}	P _{ex}
RmBra20	22	10	3	0.82	0.59	0.29	23	6	2	0.76	0.52	0.65	23	10	4	0.78	0.52	0.34	0.62
RmBra25	21	6	3	0.59	0.29	0.52*	20	7	4	0.61	0.25	0.80	15	1	0	0.0	0.00	0.00	1.00
RmBra27	20	4	1	0.27	0.50	0.82*	24	4	1	0.50	0.42	0.89	15	5	4	0.36	0.13	0.64*	0.93
RmBra45	22	10	6	0.66	0.73	-0.09	24	3	1	0.12	0.04	0.99	19	7	2	0.54	0.21	0.61*	0.84
RmBra50	24	3	0	0.29	0.17	0.44*	22	10	8	0.84	0.32	0.52	17	5	2	0.37	0.18	0.53*	0.93
RmBra59	20	5	3	0.60	0.65	-0.08	22	6	2	0.72	0.64	0.69	21	8	4	0.65	0.38	0.42	0.76
RmBra66	24	11	7	0.83	0.75	0.09	23	3	1	0.67	0.39	0.78	20	6	2	0.43	0.25	0.43	0.90
Average			7	0.58	0.46	0.21		5.6		0.60	0.31	0.48		6		0.48	0.24	0.47	

Note: A = number of alleles; A_p = number of private alleles; F_{IS} = fixation index; H_e = expected heterozygosity; H_o = observed heterozygosity; N = number of samples; P_{ex} = first parental exclusion probability.

* Indicates signification deviations of Hardy–Weinberg equilibrium.

coast revealed by microsatellite analysis. *American Journal of Botany* 98: 1031–1039.

ROSETO-GALINDO, C., E. GAITAN-SOLIS, H. CÁRDENAS-HENAO, J. TOHME, AND N. TORO-PÉREA. 2002. Polymorphic microsatellites in a mangrove species, *Rhizophora mangle* (L.) (Rhizophoraceae). *Molecular Ecology Notes* 2: 281–283.

ROZEN, S., AND H. J. SKALETSKY. 2000. Primer3 on the WWW for general users and for biologist programmers. In S. Misener and S. A. Krawetz [eds.], *Methods in molecular biology*, vol. 132: Bioinformatics methods and protocols, 365–386. Humana Press, Totowa, New Jersey, USA.

SANDOVAL-CASTRO, E., R. MUÑOZ-SALAZAR, L. M. ENRÍQUEZ-PAREDES, R. RIOSMENA-RODRÍGUEZ, R. S. DODD, C. TOVILLA-HERNÁNDEZ, AND M. C. ARREDONDO-GARCÍA. 2012. Genetic population structure of red mangrove (*Rhizophora mangle* L.) along the northwestern coast of Mexico. *Aquatic Botany* 99: 20–26.

TAKAYAMA, K., M. TAMURA, Y. TATEISHI, AND T. KAJITA. 2008. Isolation and characterization of microsatellite loci in the red mangrove *Rhizophora mangle* (Rhizophoraceae) and its related species. *Conservation Genetics* 9: 1323–1325.

TOMLINSON, P. B. 1986. *The botany of mangroves*. Cambridge University Press, Cambridge, United Kingdom.

VAN OOSTERHOUT, C., W. F. HUTCHINSON, D. P. M. WILLS, AND P. SHIPLEY. 2004. MICRO-CHECKER: Software for identifying and correcting genotyping errors in microsatellite data. *Molecular Ecology Notes* 4: 535–538.

VINSON, C. C., V. C. R. AZEVEDO, I. SAMPAIO, AND A. Y. CIAMPLI. 2005. Development of microsatellite markers for *Carapa guianensis*, a tree species from the Amazon forest. *Molecular Ecology Notes* 5: 33–34.