



Draft Genome Sequences of Six *Actinobacillus pleuropneumoniae* Serotype 8 Brazilian Clinical Isolates: Insight into New Applications

Monalessa Fábia Pereira,^a Ciro César Rossi,^a Fabíola Marques de Carvalho,^b Luiz Gonzaga Paula de Almeida,^b Rangel Celso Souza,^b Ana Tereza Ribeiro de Vasconcelos,^b Denise Mara Soares Bazzolli^a

Departamento de Microbiologia, Laboratório de Genética Molecular de Micro-organismos, Instituto de Biotecnologia Aplicada à Agropecuária–BIOAGRO, Universidade Federal de Viçosa- UFV, Viçosa, Minas Gerais, Brazil^a; Laboratório de Bioinformática, Laboratório Nacional de Computação Científica, LNCC, Petrópolis, Rio de Janeiro, Brazil^b

Actinobacillus pleuropneumoniae is the causative agent of swine pleuropneumonia, a highly contagious disease associated with pigs of all ages that results in severe economic losses to the industry. Here, we report for the first time six genome sequences of *A. pleuropneumoniae* clinical isolates of serotype 8, found worldwide.

Received 29 December 2014 Accepted 16 January 2015 Published 5 March 2015

Citation Pereira MF, Rossi CC, de Carvalho FM, de Almeida LGP, Souza RC, de Vasconcelos ATR, Bazzolli DMS. 2015. Draft genome sequences of six *Actinobacillus pleuropneumoniae* serotype 8 Brazilian clinical isolates: insight into new applications. Genome Announc 3(2):e01585-14. doi:10.1128/genomeA.01585-14.

Copyright © 2015 Pereira et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Denise Mara Soares Bazzolli, dbazzolli@ufv.br

A ctinobacillus pleuropneumoniae is the causative agent of porcine pleuropneumonia, a costly, severe, and highly contagious infectious disease (1). There are 15 known serotypes of this microorganism, the prevalence of which varies greatly worldwide (2). Serotype 8 isolates, the most widespread in southeastern Brazil (3), display high cytotoxic activity and are generally responsible for low mortality but high morbidity. This serotype is also dominant in other pig-producing countries, such as Mexico and the United Kingdom (4, 5).

We sequenced six serotype 8 A. pleuropneumoniae clinical isolates that are persistent in farms in southeastern Brazil and present different virulence degrees (6). Each isolate was grown in culture medium (BHI, BD Biosciences), and total genomic DNA was extracted using the FastDNA Spin kit (MP Biomedicals). Wholegenome sequencing was performed using an Ion Torrent personal genome machine sequencer (Life Technologies) using 200-bp chemistry with an Ion 318C chip. Assembly for each isolate was performed with Newbler version 2.9. Contigs were reordered with Mauve using A. pleuropneumoniae JL03 as the reference genome (GenBank accession no. CP000687). Automatic annotation of genes predicted with the GeneMark and Glimmer programs was performed by BLASTp against the NCBI NR, KEGG, UniProt, and TCDB databases using 100% query and subject coverage and a minimum of 95% positive as the cutoff. Manual annotation was performed using the System for Automated Bacterial Integrated Annotation (SABIA) (7).

Genome sequencing of *A. pleuropneumoniae* isolates 460, 518, 597, 780, 1,022, and 5,651 resulted in high-coverage assemblies of the 2.24 \pm 0.03 Mbp genomes (between 90- and 123-fold), with 40.33% \pm 0.31% GC content and an average coding gene length of 844.97 \pm 9.26 bp. The coding regions correspond to 82.41% \pm 0.58% of the genome, which should represent most of the functionally annotated genes and allow for comparative studies using these sequences. Approximately 2,358.16 \pm 42.01 loci were predicted for each genome, and these showed significant hits with

databases of functional data: KEGG (90.50%), UniProt (79.40%), NCBI NR (93.03%), and TCDB (23.55%). Gene prediction revealed 48.17 \pm 1.94 tRNA genes and four rRNA operons. According to a Bidirectional Best Hit comparison, the genomes present 2,296 \pm 33.32 clusters in common with strains JL03 and L20 (GenBank accession no. CP000569) and 79.5 \pm 24.05 nonclustered proteins, revealing high genetic variability of this species, often associated with intrinsic and environmental factors, such as increased horizontal gene transfer frequency, because of the remarkable ability of *A. pleuropneumoniae* to transform naturally (8, 9).

These new six *A. pleuropneumoniae* genome assemblies are the first for serotype 8 to be published in the major sequence databases. Because of the relevance of this serotype in many countries, the public availability of these sequences will be globally helpful for comparative genomic studies, thereby contributing to the discovery and identification of common virulence determinants and molecular markers that could be used for effective diagnosis and vaccine production.

ACKNOWLEDGMENTS

We thank FAPEMIG (grant APQ-00232-13), CNPq, and CAPES.

REFERENCES

- 1. Gottschalk M, Taylor DJ. 2006. *Actinobacillus pleuropneumoniae*, p 563–576. *In* Straw BE, Zimmerman JJ, D'Allaire S, Taylor DJ (ed), Diseases of swine. Blackwell Publishing Professional, Ames, IA.
- Inzana TJ, Champion A. 2007. Use of an inhibition enzyme-linked immunosorbent assay for quantification of capsular polysaccharide or proteins in vaccines. Clin Vaccine Immunol 14:323–327. http://dx.doi.org/10.1128/ CVI.00302-06.

- 3. Rossi CC, Vicente AM, Guimarães WV, Araújo EF, Queiroz MV, Bazzolli DMS. 2013. Face to face with *Actinobacillus pleuropneumoniae*: landscape of the distribution of clinical isolates in southeastern Brazil. Afr J Microbiol Res 7:2916-2924. http://dx.doi.org/10.5897/ AJMR12.2344.
- Blackall PJ, Klaasen HL, van den Bosch H, Kuhnert P, Frey J. 2002. Proposal of a new serovar of *Actinobacillus pleuropneumoniae*: serovar 15. Vet Microbiol 84:47–52. http://dx.doi.org/10.1016/S0378-1135(01)00428-X.
- O'Neill C, Jones SC, Bossé JT, Watson CM, Williamson SM, Rycroft AN, Kroll JS, Hartley HM, Langford PR. 2010. Prevalence of *Actinobacillus pleuropneumoniae* serovars in England and Wales. Vet Rec 167:661-662. http://dx.doi.org/10.1136/vr.c5106.
- 6. Pereira MF, Rossi CC, Queiroz MV, Martins GF, Isaac C, Bossé JT, Li Y, Wren BW, Terra VS, Cuccui J, Langford PR, Bazzolli DMS. 2014.

Galleria mellonella is an effective model to study *Actinobacillus pleuropneu-moniae* infection. Microbiology 161:387–400. http://dx.doi.org/10.1099/mic.0.083923-0.

- Almeida LG, Paixão R, Souza RC, Costa GC, Barrientos FJ, Santos MT, Almeida DF, Vasconcelos AT. 2004. A system for automated bacterial (genome) integrated annotation—SABIA. Bioinformatics 20:2832–2833. http://dx.doi.org/10.1093/bioinformatics/bth273.
- Bossé JT, Nash JH, Kroll JS, Langford PR. 2004. Harnessing natural transformation in *Actinobacillus pleuropneumoniae*: a simple method for allelic replacements. FEMS Microbiol Lett 233:277–281. http://dx.doi.org/ 10.1016/j.femsle.2004.02.022.
- Xu Z, Chen X, Li L, Li T, Wang S, Chen H, Zhou R. 2010. Comparative genomic characterization of *Actinobacillus pleuropneumoniae*. J Bacteriol 192:5625–5636. http://dx.doi.org/10.1128/JB.00535-10.